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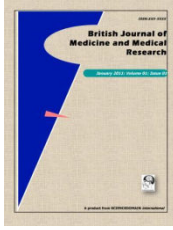
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A Mediterranean Low-Glycemic-Load Diet alone or in Combination with a Medical Food Improves Insulin Sensitivity and Reduces Inflammation in Women with Metabolic Syndrome

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Research Article

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ABSTRACT

Aim: To determine the effects of a Mediterranean-style low-glycemic load diet alone or in combination with a medical food (MF) on insulin resistance and inflammation in women with metabolic syndrome (MetS).

Study design: Two groups, Parallel study with control.

Place and Duration of Study: Department of Nutritional Sciences, University of Connecticut, Storrs, CT; Department of Emergency Medicine, University of Florida, Jacksonville, FL; Department of Medicine, University of California, Irvine, CA. The study was carried out from September of 2009 to May 2010.

Methodology: Eighty three women (20-75 y) with MetS. Participants were randomly allocated to consume diet alone (control group) or the diet plus the MF (MF group) for 12 wk. Body composition was measured at baseline, week 8 and week 12 by use of bioelectrical impedance in all participants while Dual-emission X-ray absorptiometry was

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used for 37 of the subjects. Insulin resistance, plasma insulin, leptin, adiponectin and the inflammatory cytokines, tumor necrosis factor (TNF)- α , interleukin-6 (IL-6), adhesion molecules, sICAM-1 and sVCAM-1, were measured at the same time points.

Results: Independent of group allocation, women had decreases in body mass index ($p < 0.0001$) and body and trunk fat ($p < 0.0001$). Plasma insulin, insulin resistance, and leptin were also significantly decreased over time ($p < 0.0001$), while plasma adiponectin levels did not change. Regarding inflammatory markers, significant reductions were found in TNF- α ($p < 0.0001$) and sICAM-1 levels ($p < 0.001$), but not in IL-6 or sVCAM-1. At 12 wk, sICAM was reduced only in the MF group ($p < 0.05$ vs control).

Conclusion: A Mediterranean-style diet, in combination with a MF, are a viable option to decrease factors associated with increased risk for type 2 diabetes and coronary heart disease.

Keywords: Metabolic syndrome; mediterranean diet; glycemic load; insulin; inflammation; body composition;

This trial was registered at clinicaltrials.gov as NCT01010841.

ABBREVIATIONS

BMI: body mass index; BP: blood pressure; CVD: cardiovascular disease; DEXA: dual-energy X-ray absorptiometry; HDL-C: HDL cholesterol; ICAM: Intracellular adhesion molecule; IL-6: interleukin-6; LDL-C: LDL cholesterol; MetS: Metabolic syndrome; NHANES: National Health and Nutrition Examination Survey; T2D: type 2 diabetes; TNF- α : tumor necrosis alpha; VCAM: vascular cell adhesion molecule-1 WC: waist circumference; WC: waist circumference.

1. INTRODUCTION

Insulin resistance, the diminished capacity of a cell to respond to insulin action, has been established as a primary underlying cause of metabolic syndrome (MetS) development (Lann, Gallagher et al. 2008; Gallagher, Leroith et al., 2010; Maheshwari and Thuluvath, 2011). An individual is classified as having MetS if 3 of the following criterion are met: hyperglycemia, high waist circumference (WC), elevated blood pressure (BP), elevated triglycerides (TG), and low high-density lipoprotein cholesterol (HDL-C) (Alberti, Eckel et al. 2009). According to the National Health and Nutrition Examination Survey (NHANES) 2003-2006, approximately one-third of the US adult population is affected by MetS, and this significantly increases the risk for developing cardiovascular disease (CVD) and type 2 diabetes (T2D) (Ervin 2009).

While insulin resistance is often considered the causal feature for development of MetS, abdominal obesity is also a major contributor and the two components are clearly related (Després, 2006; Després, Lemieux et al., 2008; Westphal, 2008; Reddy, Singh et al., 2010). Adipose tissue is considered an active endocrine organ (Wozniak, Gee et al., 2009; Han and Lean, 2011), responsible for secreting biologically active molecules, collectively termed 'adipokines' (Caruso, Balistreri et al., 2010). When adipose tissue is excessive, secretion of

adipokines such as adiponectin, leptin, interleukin 6 (IL-6) and tumor necrosis factor alpha (TNF- α) can be negatively altered (Lago, Dieguez et al., 2007; Lago, Gómez et al., 2009), which can in turn influence energy regulation and inflammation (Han and Lean, 2011). Adiponectin, an insulin sensitizer with anti-inflammatory properties, tends to be decreased in obesity and MetS (Ryo, Nakamura et al. 2004; Huang 2009). Leptin is important for regulation of appetite and energy balance. In obese individuals, despite elevated leptin concentrations, sensitivity to this molecule has been shown to decrease (Gnacińska, Małgorzewicz et al., 2009). Proinflammatory adipokines, IL-6 and TNF- α , also tend to be upregulated in obesity, and these molecules have a role in insulin resistance and endothelial dysfunction (Ryo, Nakamura et al. 2004; Maury and Brichard 2010; Van Linthout, Foryst-Ludwig et al. 2010). These inflammatory mediators alter insulin sensitivity via action on nuclear factor kappa B (NF κ B) and jun N-terminal kinase (JNK) pathways, and they facilitate expression of intercellular adhesion molecule-1 (ICAM-1) and vascular cell adhesion molecule-1 (VCAM-1), which both play a role in atherogenesis (Lyon, Law et al., 2003; Shoelson, Herrero et al., 2007). Obesity has also been associated with macrophage accumulation in adipose tissue, and this could further contribute to the production of proinflammatory cytokines (Weisberg, McCann et al., 2003).

The central focus to treat inflammation, excess abdominal fat stores, and insulin resistance that compose MetS, remains lifestyle modification. Various researchers have highlighted the potential efficacy of adherence to a Mediterranean-style diet for treatment of MetS characteristics (Babio, Bulló et al., 2009; Sofi, 2009; Kastorini, Milionis et al., 2011). Epidemiological data suggest that prevalence of MetS tends to be associated with dietary patterns that contain excessive amounts of high glycemic index foods (Baxter, Coyne et al. 2006). Therefore, our aim was to examine the impact of a low-glycemic-load Mediterranean-style diet on specific MetS variables, namely inflammation, insulin resistance, and obesity. Additionally, our objective was to assess whether a medical food (MF), containing *rho* iso-alpha acids from hops (*Humulus lupulus* L.) and proanthocyanidins from acacia (*Acacia nilotica*), which may modulate insulin-signaling and inflammatory pathways (Lerman, Minich et al., 2008), provided further benefits. We hypothesized that following this dietary intervention would result in similar weight loss for all participants, but the group consuming the MF would have greater improvements in insulin sensitivity and present a better inflammatory profile.

2. METHODS

2.1 Materials

The medical food (UltraMeal[®] Plus 360[°]) was provided by Metagenics, Inc (Gig Harbor, WA), in powdered form. Kits to measure adiponectin, sICAM, sVCAM, TNF- α and IL-6 were obtained from Millipore (Billerica, MA).

2.2 Study Population

Women were recruited at three sites (University of Connecticut, Storrs, CT; University of Florida, Jacksonville, FL and University of California, Irvine, CA). To be eligible for participation, women had to be between the ages of 20 and 75 years with a BMI between 25 and 45 kg/m². In addition, they had to have LDL-C \geq 100 mg/dL (2.59 mmol/L), TG \geq 150 mg/dL (1.70 mmol/L), and meet two of the four remaining criteria for MetS according to NCEP-ATP-III criteria [HDL-C < 50 mg/dL (1.30 mmol/L), fasting plasma glucose \geq 100

mg/dL (5.55 mmol/L), blood pressure (BP) \geq 130/85 mm Hg, and WC > 88 cm) (ATP, (2002). Main exclusion criteria included heart, liver, or kidney disease and use of hypoglycemic or cholesterol-lowering agents. Potential participants provided informed consent during screening and before the study began. The women also provided a medical history prior to participation. Study protocols and the procedures to obtain consent forms were approved by the Institutional Review Board at each respective site.

2.3 Study Design

After entrance criteria were met and consent was obtained, women were randomly allocated to the control group or the medical food (MF) group. Both groups were given detailed instructions to consume a modified Mediterranean-style, low-glycemic-load diet for the duration of the 12-week intervention. Women in the MF group consumed the same diet plus a powdered beverage form of a medical food containing soy protein, plant sterols, *rho* iso-alpha acids, and *Acacia* proanthocyanidins, mixed into water, twice each day for 12 weeks. Participants came on-site for a baseline visit and returned at weeks 2, 4, 6, 8, 10, and 12 for follow-up visits. Compliance with MF intake guideline was assessed by weighing the unused powder every 2 weeks.

2.4 Diet and Exercise Description

Participants were invited to an on-site meeting 1 week prior to the start of the study, at which time an instructional DVD was viewed and detailed dietary booklets were distributed summarizing dietary guidelines, lists of suitable foods, suggested serving sizes, and recipes. Subjects were also instructed on how to complete a detailed food record that included 2 weekdays and one weekend day. The dietary approach was parallel to that designed by Schiltz et al. (Schiltz, Minich et al., 2009). Subjects were advised to follow a Mediterranean-style diet with emphasis on intake of legumes, fish and lean meats. In addition, specific limitations were placed on intake of grains (1 serving per day) and alcohol (an optional 1 serving per day). Lastly, natural and artificial sweeteners were excluded from the diet. The women were not instructed to restrict caloric intake. A 3-day food record was obtained at baseline (to assess habitual dietary intake) and every 2 weeks (to assess dietary compliance and nutrient intake). The participants met with a Registered Dietitian at each visit for diet counseling. Subjects were asked to maintain their normal level of physical activity for the entirety of the intervention and 7-day exercise logs were collected to measure compliance with this instruction.

2.5 Data Collection

Following a 12-hour overnight fast, 60 mL of blood were collected into EDTA tubes at baseline, week 8, and week 12. Plasma was separated by centrifugation at 2000 x g: 20 min at 4°C. The plasma was aliquoted to cryotubes and stored at -80 °C and later sent to a certified laboratory (Northwest Lipid Research Laboratories, Seattle, WA) for analysis of insulin, or used at the University of Connecticut for analysis of adipokines and inflammatory markers.

2.6 BMI and Body Composition

At each visit, body weight was measured to the nearest 0.1 kg on a calibrated digital scale, and BMI was calculated (kg/m^2). At baseline, week 8 and week 12, bioelectrical impedance

analysis (BIA) was used to assess body composition, utilizing an Impedimed DF 50 (Xitron Technologies, San Diego, CA) at all sites. In addition to the BIA, the University of Connecticut and the University of California, Irvine, determined whole body and regional body composition using fan-beam dual-energy X-ray absorptiometry (DEXA) (Prodigy™, Lunar Advance Corporation, Madison, WI). Analyses were performed by a technician at the respective sites (MF: n = 18, control: n = 17).

2.7 Plasma Insulin and Insulin Resistance

Plasma insulin was measured by a chemiluminescent, immunoassay using the DPC Immulite 2000 (Diagnostics Products Corporation, Nutley, NJ) at Northwest Lipid Research Laboratories (Seattle, WA). Insulin resistance was calculated using the homeostasis model assessment (HOMA) equation ($[\text{fasting insulin } (\mu\text{U/ml}) \times \text{fasting glucose (mmol/l)}] / 22.5$) (Muniyappa, Lee et al., 2008).

2.8 Fasting Adiponectin, sVCAM-1, and sICAM-1

Fasting plasma was used to measure concentrations of adiponectin, sVCAM-1, and sICAM-1 in duplicate using the Milliplex Human CVD Panel 1 (City, State). The samples were diluted 1:100 and antibody-immobilized beads specific for each analyte were used for simultaneous quantification with xMAP® technology on the Luminex® IS 200 system (Austin, TX).

2.9 Fasting IL-6, TNF- α , and Leptin

Fasting plasma was used to measure concentrations of IL-6, TNF- α , and leptin in duplicate using the Milliplex Human Cytokine kit and xMAP® technology on the Luminex® IS 200 system (Austin, TX). Specific antibody-immobilized beads were used to simultaneously quantify IL-6, TNF- α , and leptin.

2.10 Statistical Analysis

Repeated measures analysis of variance (ANOVA) was used to compare the control and MF groups over time. The repeated measure was the individual's response to the intervention over time and the control vs. the MF served as the between subject factors. SPSS 12.0 for Windows (SPSS, Inc., Evanston, IL) was used and results were considered significant with a $p < 0.05$.

3. RESULTS

3.1 BMI and Body Composition

Participants in both groups had significant decreases in BMI from baseline to the end of the 12-week intervention ($p < 0.0001$, Table 1), but no differences were observed between the groups. Examination of BIA measurements revealed that fat mass, in kg and as a percentage of total body weight, decreased significantly over time ($p < 0.0001$ Table 1), regardless of group categorization. Likewise, fat free mass in kg decreased in both groups ($p < 0.0001$) while percent of fat free mass increased also for both groups ($p < 0.001$, Table 1), with no differences noted between groups.

Table 1. Body mass index (BMI) and body composition as measured by bioelectrical impedance analysis of women allocated to the Medical Food group (n = 42) or the Control group (n = 41) at baseline, week 8 and week 12 of the intervention¹

	BMI kg/m ²	Fat Free Mass kg	Fat Mass	Fat Free Mass %	Fat Mass
Medical Food Group					
Baseline	34.9 ± 6.1 ^a	50.7 ± 5.7 ^a	41.8 ± 10.5 ^a	55.7 ± 4.3 ^a	44.2 ± 4.3 ^a
Week 8	33.1 ± 5.5 ^b	49.4 ± 5.4 ^b	38.4 ± 10.6 ^b	57.4 ± 5.9 ^b	42.6 ± 5.9 ^b
Week 12	32.5 ± 5.8 ^b	48.8 ± 5.2 ^b	37.7 ± 10.4 ^b	57.6 ± 5.7 ^b	42.4 ± 5.7 ^b
Control Group					
Baseline	35.0 ± 5.5 ^a	50.7 ± 5.8 ^a	39.4 ± 9.0 ^a	56.0 ± 3.6 ^a	43.9 ± 3.5 ^a
Week 8	33.1 ± 5.0 ^b	49.7 ± 5.6 ^b	36.1 ± 9.2 ^b	57.8 ± 5.4 ^b	42.2 ± 5.2 ^b
Week 12	32.7 ± 5.1 ^b	49.5 ± 5.6 ^b	35.3 ± 9.5 ^b	58.4 ± 6.1 ^b	41.7 ± 5.8 ^b
Time Effect	P < 0.0001	P < 0.0001	P < 0.0001	P < 0.0001	P < 0.001

¹Values are expressed as mean ± SD. No diet effects or interactive effects were found for these variables. Values with different superscripts are significantly different as compared to baseline.

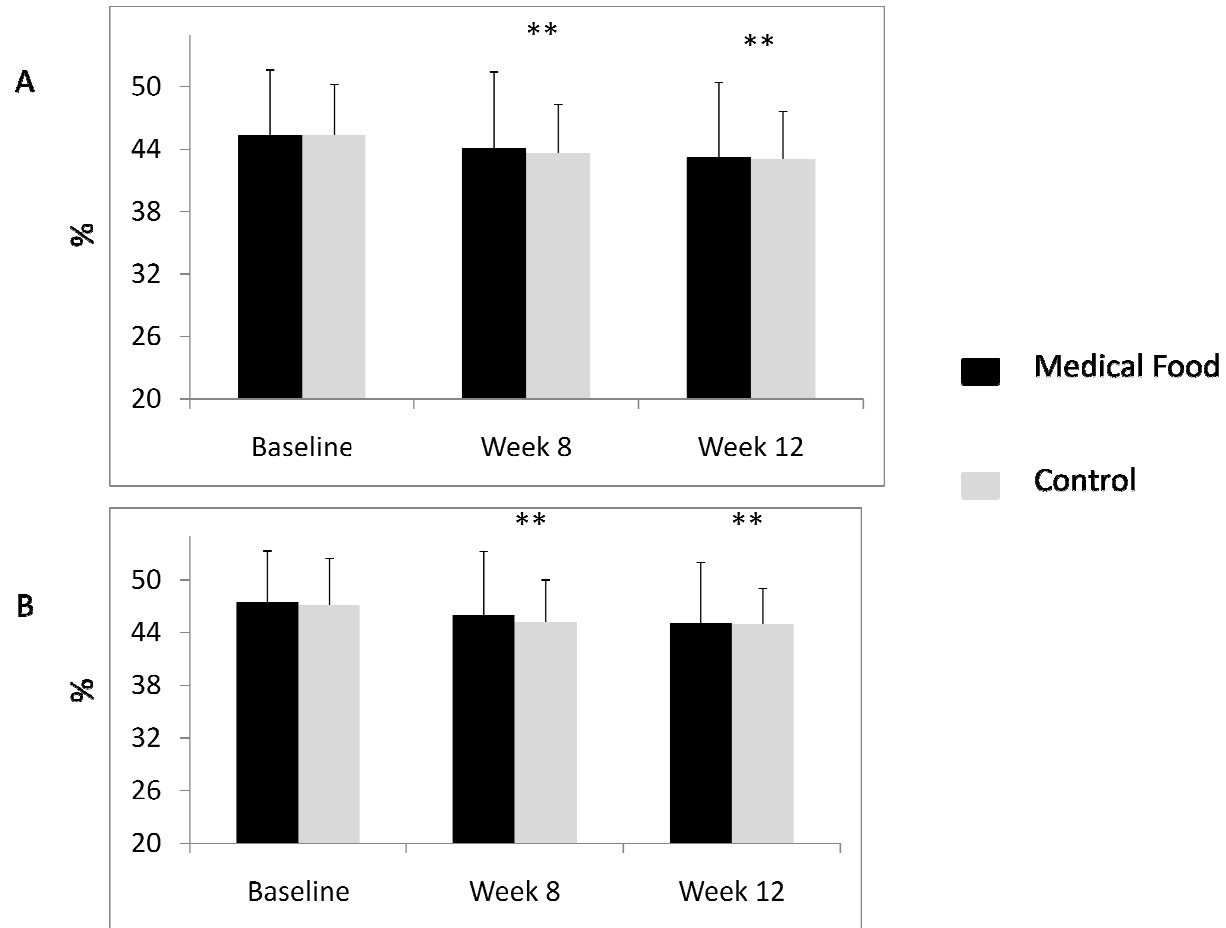


Fig. 1. Percent Body Fat (A) and Percent Trunk Fat (B) calculated by DEXA at baseline week 8 and week 12 for women allocated to Medical Food (n = 18) or Control (n = 17). ** p < 0.0001.

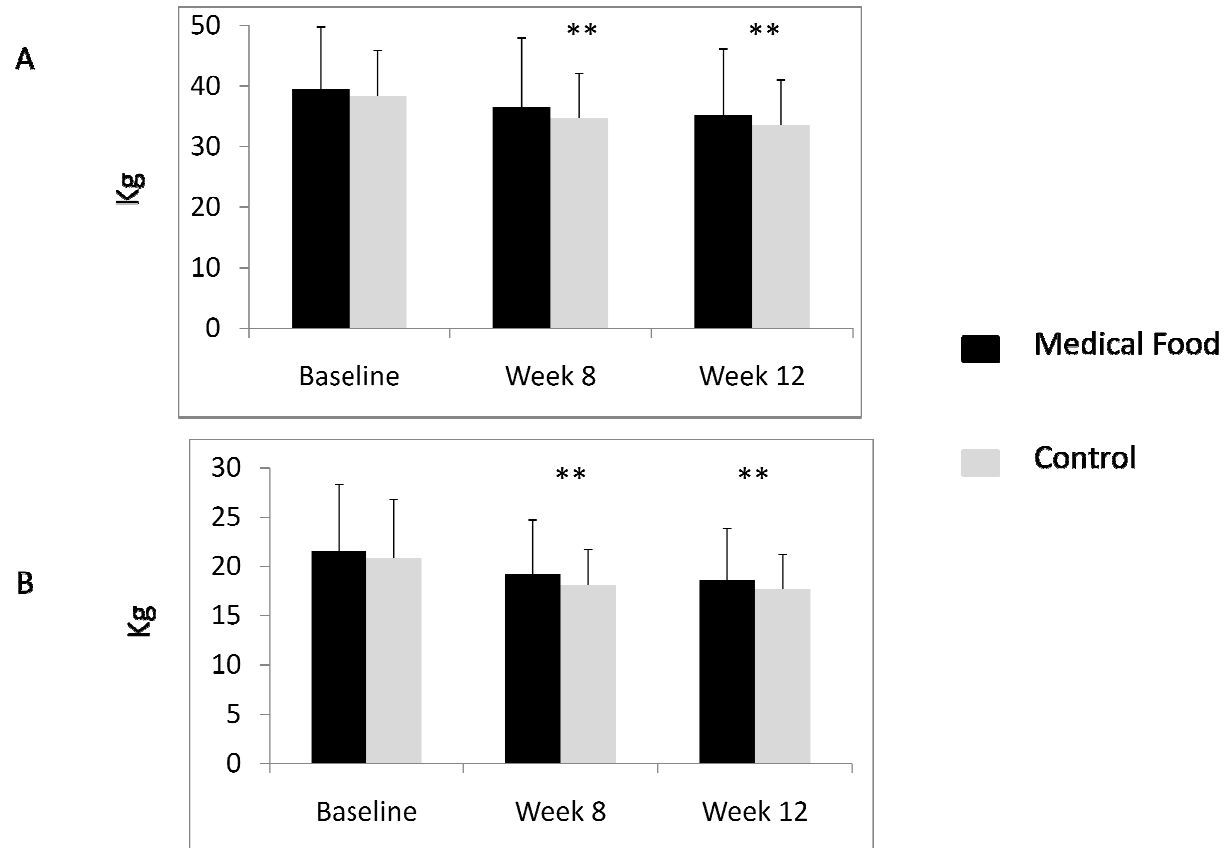


Fig. 2. Body Fat Weight (A) and Trunk Fat Weight (B) calculated by DEXA at baseline week 8 and week 12 for women allocated to Medical Food (n = 18) or Control (n = 17). ** p < 0.0001.

The University of Connecticut and the University of California, Irvine, had access to DEXA, which allowed measurements of body fat and trunk fat for a subset of subjects (MF: n = 18, control: n = 17). Percent body fat and percent trunk fat decreased in both groups, with no differences between groups ($p < 0.0001$; A and B, respectively, Figure 1).

Similarly, there were no differences between groups in relation to the significant reductions observed in kg of body fat and trunk fat ($p < 0.0001$, A and B, respectively, Figure 2).

3.2 Insulin and Insulin Resistance

Plasma insulin levels decreased in both groups over the course of the 12-week intervention, independent of group allocation ($p < 0.0001$, Table 2). HOMA scores were utilized to assess insulin resistance, and both groups displayed reductions in this variable ($p < 0.0001$, Table 2), with no between-group differences noted.

Table 2. Changes in insulin and insulin resistance as measured by HOMA of women allocated to the Medical Food group (n = 42) or the Control group (n = 41) at baseline, week 8 and week 12 of the Intervention¹

	Insulin μU/mL	HOMA*
Medical Food Group		
Baseline	16.9 ± 9.4 ^a	4.25 ± 2.93 ^a
Week 8	12.3 ± 6.0 ^b	3.01 ± 1.57 ^b
Week 12	12.6 ± 7.6 ^b	2.99 ± 1.91 ^b
Control Group		
Baseline	16.6 ± 9.4 ^a	4.12 ± 2.40 ^a
Week 8	11.8 ± 5.4 ^b	2.84 ± 1.30 ^b
Week 12	12.2 ± 5.7 ^b	3.01 ± 1.58 ^b
Time Effect	P < 0.0001	P < 0.0001

¹Values are expressed as mean ± SD. No diet effects were found for these variables. Values with different superscripts are significantly different as compared to baseline.

3.3 Adipokines and Inflammatory Markers

Plasma adiponectin levels did not change significantly in either group over time (Table 3). However, changes were observed in plasma levels of leptin ($p < 0.0001$, Table 3), although the changes were not significantly different between groups. While plasma levels of IL-6 did not change significantly in either group (Table 3), there were differences observed over time for plasma TNF-α ($p < 0.0001$, Table 3); however, these were not different between groups at the end of the intervention. sICAM-1 decreased in the MF group over time ($p < 0.001$, Table 3), while levels in the control group were not different from baseline in either week 8 or 12 (Table 3). No significant changes in sVCAM-1 were measured for either group (Table 3).

Table 3. Changes in adiponectin, tumor necrosis factor (TNF)- α , interleukin-6 (IL-6), soluble intercellular adhesion molecule-1 (ICAM)-1 and soluble vascular cell adhesion molecule (VCAM)-1 of women allocated to the Medical Food group (n = 42) or the Control group (n = 41) at baseline, week 8 and week 12 of the intervention¹

	Adiponectin mg/L	Leptin	TNF- α	IL-6	sICAM-1	sVCAM-1
Medical Food Group						
Baseline	15.2 \pm 7.9 ^a	51.0 \pm 30.1 ^a	4.6 \pm 2.0 ^a	5.1 \pm 3.4 ^a	112 \pm 55 ^a	1.3 \pm 3.4 ^a
Week 8	15.7 \pm 6.8 ^a	36.5 \pm 21.8 ^b	4.3 \pm 1.6 ^b	4.8 \pm 3.2 ^a	96 \pm 40 ^b	1.2 \pm 2.9 ^a
Week 12	15.0 \pm 10.7 ^a	34.2 \pm 23.0 ^b	4.2 \pm 1.8 ^b	4.7 \pm 3.7 ^a	96 \pm 37 ^b	1.2 \pm 2.7 ^a
Control Group						
Baseline	14.2 \pm 6.8 ^a	55.5 \pm 27.5 ^a	5.4 \pm 2.4 ^a	4.3 \pm 2.5 ^a	105 \pm 29 ^a	1.2 \pm 2.6 ^a
Week 8	13.8 \pm 6.0 ^a	41.7 \pm 25.8 ^b	5.1 \pm 2.1 ^{ab}	4.0 \pm 2.1 ^a	99 \pm 32 ^{ab}	1.2 \pm 3.0 ^a
Week 12	14.8 \pm 6.3 ^a	40.4 \pm 25.7 ^b	4.3 \pm 1.7 ^b	3.9 \pm 2.4 ^a	103 \pm 49 ^a	1.2 \pm 3.3 ^a
Time Effect	NS	P < 0.0001	P < 0.0001	NS	P < 0.001	NS
Diet Effect	NS	NS	NS	NS	P < 0.05	NS

¹Values are expressed as mean \pm SD. Values with different superscripts are significantly different as compared to baseline.

4. DISCUSSION

A consensus has not been reached regarding the best dietary pattern to alleviate characteristics of MetS, but the Mediterranean-style pattern has received ample attention and is considered an effective option (Djoussé, Padilla et al., 2010). The aim of the current study was to evaluate the impact of a Mediterranean-style, low-glycemic-load diet on body composition, insulin resistance and inflammation, in a population of women with MetS and elevated LDL-C. Another objective was to determine if a phytosterol-rich MF provided any additional benefits to these parameters. Following the 12- week intervention, both groups showed improvements in body composition, insulin resistance and inflammatory profiles.

Generally, a positive correlation exists between BMI, adiposity and risk for T2D and CVD; therefore, BMI is typically considered a suitable tool to define adult obesity ($BMI \geq 30 \text{ kg/m}^2$) and pinpoint individuals that may be at risk of obesity-related morbidity and mortality (2003; Westphal 2008). BMI is often used as a surrogate marker of overall obesity, while WC is used to assess central obesity. Both of these measurements are easily obtainable indirect measurements of obesity (Koopman, Swofford et al., 2009). WC and BMI are strongly correlated, and elevated WC alone is a strong predictor of mortality and risk for chronic disease development (Phillips and Prins, 2008; Achike, To et al., 2011). Measurements of WC are better indicators of body fat distribution, and it is not total body fat, or subcutaneous fat, but rather, excess intra-abdominal fat that is highly associated with metabolic abnormalities in overweight and obese individuals (Bjorntorp, 1991; Nielsen and Jensen, 1997; Després, 2006). We previously reported significant reductions in weight and WC for the participants of this study (Jones, Fernandez et al., 2011), and here we show that BMI was significantly reduced in both groups post-intervention. While BMI is a standard indirect measure of overweight and obesity, it does not distinguish between fat mass and fat free mass, nor does it describe body fat distribution (Boardley and Pobocik, 2009).

In order to provide a more comprehensive account of changes in body composition over the course of the intervention, BIA and DEXA measurements were utilized. Results from the BIA showed that all participants had significant reductions in absolute (kg) and relative (%) amounts of fat mass while percent free fat mass increased after the intervention. Results from DEXA assessment revealed significant decreases in body fat and trunk fat, on a percentage and weight basis. While the decreases in fat mass are desirable, those observed in fat free mass (specifically lean mass) are less so, but it is acknowledged that body weight decreased and percent lean increased over time. Abdominal obesity has been considered a dominant cause of MetS (Després, 2006); therefore, the specific losses in trunk fat may decrease cardiometabolic risk (Katsoulis, Blaudeau et al., 2009).

Weight loss, and especially abdominal fat loss, has been associated with improvements in insulin resistance (Weiss, 2007; Bellar, Jarosz, et al., 2008; Aasen, Fagertun et al., 2010). In a previous study of similar design to the current one, supplementation with a MF and identical phytochemical-combination increased insulin sensitivity (Lerman, Minich et al., 2008), it was the expectation that the same observation would be made following this intervention. While we did measure significant reductions in insulin, and improvements in HOMA-IR scores, the changes were not statistically different between groups. Insulin resistance is associated with CVD (Ferrannini, Balkau et al., 2007), and if ignored, can progress to T2D (Phillips and Prins, 2008). In conjunction with improvements in insulin resistance, we expected to observe increases in plasma adiponectin, which has insulin-sensitizing and anti-inflammatory properties (Matsuzawa, 2010). However, we did not obtain any significant changes in adiponectin levels for either group. Polak and Shin also reported

no changes in adiponectin levels, despite weight loss in women (Shin, Yoen Kim, et al., 2006; Polak, Kovacova et al., 2008). This suggests that other weight loss-associated factors, in organs other than adipose, may account for the improved insulin variables.

Despite the lack of weight-loss induced change on adiponectin, leptin levels did decrease significantly in both groups over time. Leptin, an adipokine responsible for increasing energy expenditure and suppressing food intake (Matsuzawa, 2005), tends to be elevated in obese individuals, indicative of leptin resistance (Gade, Schmit et al., 2010; Münzberg, 2010). Therefore, the decreases observed may suggest improved sensitivity to leptin. The significant decreases observed in trunk fat may also contribute to the reductions in plasma levels of TNF- α . In an obese state, adipocytes can overproduce cytokines such as TNF- α and IL-6 (Jensen, 2006). These molecules in turn recruit monocytes to the adipose tissue and facilitate their differentiation to active macrophages, which can produce more pro-inflammatory molecules, intensifying the systemic metabolic dysfunction associated with obesity (Ouchi, Parker et al., 2011; Vykoukal and Davies, 2011). We did not observe substantial changes in IL-6, but the reductions in TNF- α are indicative of a less inflammatory profile. Another marker of inflammation, and more specifically of endothelial function, sICAM-1 was also reduced, but only in the MF group. ICAM-1 responds to inflammatory conditions and mediates the migration of leukocytes into tissues (Hubbard and Rothlein, 2000). There is a tendency for ICAM-1 to be elevated in obesity, and it has been positively correlated with central adiposity and insulin resistance (Strackowski, Lewczuk et al., 2002; Leinonen, Hurt-Camejo et al., 2003). In agreement with the similar reductions we recorded in abdominal fat and insulin resistance for both groups, the decreases in sICAM-1 were expected regardless of group distribution. However, we only observed changes in the MF for this variable. VCAM-1, another marker of endothelial function was not significantly modified in either group following the intervention.

We acknowledge the limitation of only utilizing DEXA, the gold-standard to assess body composition, for a subset of study participants. It may also have been beneficial to assess adiponectin levels using a more sensitive assay, such as an ELISA, but sample availability made this unfeasible.

5. CONCLUSIONS

Overall, following the 12-week intervention, BMI decreased significantly in both groups, but not enough on average to remove participants from the obese category ($BMI \geq 30 \text{ kg/m}^2$). However, direct analysis of body composition, with BIA and DEXA, revealed significant decreases in abdominal fat mass. These data, in combination with the previously reported WC information (Jones, Fernandez et al., 2011), substantiate the possible benefits of a Mediterranean-style dietary pattern for obesity treatment, and more specifically for visceral adipose reduction. Furthermore, insulin resistance and leptin sensitivity were improved in all participants. The MF appeared to improve endothelial function, as evidenced by the reductions in sICAM-1, which were not observed in the control group. In total, adherence to a low-glycemic-load, Mediterranean-style diet appears to positively influence body composition and insulin resistance in women with MetS and elevated LDL-C.

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